Utility of Treadmill Testing in Identification and Genotype Prediction in Long-QT Syndrome

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Background—The clinical diagnosis of long-QT syndrome (LQTS) remains challenging when ECG abnormalities are borderline or intermittent. Despite issues with access, cost, and heterogeneity of LQTS mutations, genetic testing remains the diagnostic gold standard for diagnosis of LQTS. We sought to develop a provocative testing strategy to unmask the LQTS phenotype and relate this to the results of genetic testing.

Methods and Results—From 1995 to 2008, 159 consecutive patients with suspected LQTS underwent provocative testing that consisted of a modified Bruce protocol treadmill exercise test, with ECGs recorded supine at rest, immediately on standing, and at 1-minute intervals during exercise, at peak exercise, and at 1-minute intervals during the recovery phase. Similar testing was carried out on a stationary bike in a gradual and burst exercise fashion. LQTS was confirmed with genotyping in all 95 affected LQTS patients and excluded with negative family screening in 64 control subjects. Patients were studied before and after initiation of β-blockers. Of 159 patients, 50 had an LQT1 mutation and 45 had an LQT2 mutation. In the LQTS group, 44.3% of patients had a normal-to-borderline resting QTc interval. LQTS patients exhibited a greater prolongation in QTc with postural change than unaffected patients (LQT1: 40 ms [IQR, 42]; LQT2: 35 ms [IQR, 46]; and LQTS-negative: 21 ms [IQR, 37]; P = 0.029). During exercise, LQT1 patients had marked QTc prolongation compared with LQT2 and LQTS-negative patients (LQT1: 65 ms [60], LQT2: 3 ms [46], LQTS negative: 5 ms [41]; P < 0.0001). QT hysteresis was more pronounced in patients with LQT2 mutations compared with LQT1 and LQT-negative patients (LQT2: 40 ms [10], LQT1: 15 ms [40]; LQTS-negative: 20 ms [20]; P < 0.001). β-Blockade normalized the QTc changes seen with standing and QT hysteresis.

Conclusions—The presence and genotype of LQTS can be predicted by a combination of postural and exercise changes in the QT/RR relationship. β-Blockade normalized these changes. Routine exercise testing is useful in predicting and directing genetic testing in LQTS. (Circ Arrhythm Electrophysiol. 2010;3:120-125.)

Key Words: exercise ■ QT interval ■ long-QT syndrome ■ diagnosis

Long-QT syndrome (LQTS) is a cardiac channelopathy that is characterized by a prolonged QT interval, syncope, ventricular arrhythmias, and sudden death. Long-QT mutations are heterogeneous in their phenotypic expression. The great majority (>90%) of mutations involve the LQT1 (KCNQ1) or LQT2 (KCNH2) genes, which encode for the Iκs and Ikᵣ potassium channels, respectively. In the United States, it is estimated that 4000 people die of resultant sudden cardiac death yearly, the majority in childhood or adolescence. Excellent outcomes in the management of LQTS have been achieved by lifestyle modification, β-blocker therapy, and selective ICD implantation.

Clinical Perspective on p 125

The diagnosis of LQTS remains challenging in patients with borderline ECG abnormalities, and a prolonged QT interval is often overlooked. Genetic testing has come to the forefront as a powerful tool to identify patients with LQTS. Yet, it remains expensive and unavailable to many centers. Furthermore, up to 50% of patients with LQTS can have a normal-to-borderline prolonged QT interval ("concealed" LQTS), making selection of patients for genetic testing difficult. Finally, genetic testing may identify novel LQTS mutations of unclear significance, which could represent normal variants (false-positives, single nucleotide polymorphisms), and require validation. A provocative testing strategy is needed to help unmask the LQTS phenotype to not only guide genetic testing and assist with the diagnosis of LQTS but also to validate the significance of genetic findings. We examined the utility of a provocative postural and exercise test protocol in identifying patients with LQT1 and LQT2 mutations among patients referred for evaluation of LQTS at a regional Inherited Arrhythmia Clinic.

Methods

Patients with suspected LQTS were referred to the Inherited Arrhythmia Clinic at the London Health Sciences Centre for assessment.
Inclusion criteria included a history of syncope or cardiac arrest and either the presence of an affected first-degree relative or a borderline-to-prolonged corrected QT interval ( \( \geq 440 \text{ ms in men and } \geq 460 \text{ ms in women} \) on their resting 12-lead ECG. Probandts underwent comprehensive direct sequencing of the complete KCNQ1, KCNH2, SCN5A, KCNE1, and KCNE2 genes in either a research laboratory or through PGx Health (New Haven, Conn). Sequencing included PCR amplification, forward and reverse sequencing, and comparison with 500 to 750 normal multiethnic volunteers to detect common polymorphisms. Family members of probands with suspected disease-causing mutations underwent family-specific screening only. Consecutive patients with genetic data between 1995 and 2008 were included in the analysis. Six LQT3 patients were excluded from the study because of insufficient numbers to permit meaningful comparison. The protocol was approved by the University of Western Ontario Ethics Review Board, and all patients provided informed consent.

Provocative testing consisted of a modified Bruce protocol treadmill test and 2 forms of exercise on a stationary bicycle as described previously. In the patients with implanted devices, lower rate was temporarily programmed to as low as 40 bpm to ensure intrinsic rhythm during exercise testing. In brief, we obtained 12-lead ECGs of study patients while supine, immediately on standing, and at 1-minute intervals during exercise, at peak exercise, and at 1-minute intervals during the 6-minute recovery phase. Continuous ECGs were also obtained during burst and gradual bicycle exercise testing and during recovery. Blood pressure, heart rate, and symptoms were assessed every 1-minute intervals during exercise testing. In brief, we obtained 12-lead ECGs of study patients while supine, immediately on standing, and at 1-minute intervals during exercise, at peak exercise, and at 1-minute intervals during the 6-minute recovery phase. Continuous ECGs were also obtained during burst and gradual bicycle exercise testing and during recovery. Blood pressure, heart rate, and symptoms were monitored during testing. The QT interval was measured as the time interval in milliseconds from the beginning of the QRS complex and the end of the T-wave. The end of the T-wave was determined as the intersection point between the isoelectric baseline and the tangent line representing the maximal downward slope of the positive T-wave or maximal upward slope of the negative T-wave. The QT line representing the maximal downward slope of the positive T-wave was considered the longest interval of all 12 leads, primarily measured in lead II and V5. The mean of 3 QT intervals was used. The corrected QT (QTc) was calculated using the Bazett formula.

Continuous variables were compared by use of Mann-Whitney U tests for 2-group comparisons and Kruskal-Wallis tests for multiple group comparisons. Probability values for multiple comparisons were corrected using a Bonferroni adjustment. \( \chi^2 \) tests were used for comparison of categorical variables. Results are shown in medians and interquartile range (IQR). Continuous variables are expressed as mean±SD. Test performance was assessed by use of receiver operator characteristic (ROC) curves and presented with a probability value testing the null hypothesis that the area under the curve is 0.5. Statistical analysis was performed using SAS software version 9.1 (SAS Institute, Cary, NC) by the authors. Probability values of <0.05 were considered significant.

**Results**

**Patients**

Between 1995 and 2008, 159 patients and family members were assessed for LQTS (Table 1). Ninety-five patients were genotype-positive for LQTS, with 50 LQT1 patients (28 women; mean age, 36±19 years) and 45 LQT2 patients (34 women; mean age, 34.5±21 years). A total of 64 patients (39 women; mean age, 38±20 years) were genotype-negative, representing unaffected family members that served as the control population. The resting QTc was prolonged in patients with LQTS mutations compared with LQTS-negative patients (\( P<0.0001 \)). In the LQTS group, 24.3%, 20%, and 55.7% had a normal, borderline, and prolonged resting QTc, respectively. In the control subjects, 80.5%, 13.9%, and 5.6% had a normal, borderline, and prolonged resting QTc, respectively.

**Response of the QTc Interval to Change in Posture and Exercise**

Patients with LQTS exhibited a greater prolongation in QTc on standing than LQTS-negative patients (Figure 1A, \( P=0.029 \)). During exercise, patients with LQT1 mutations had an attenuated QT shortening compared with LQTS-negative and LQT2 patients (Figure 1B, \( P<0.0001 \)). In addition, LQT1 patients had a marked QTc prolongation during exercise, whereas only a modest change in the QTc interval was observed in LQT2 and LQTS-negative patients (Figure 1C, \( P<0.0001 \)). Burst bike and gradual bike testing showed similar results, with LQT1 patients demonstrating significantly impaired shortening of their QT interval and pronounced QTc lengthening with exercise (Figure 1B and 1C). In contrast, LQT2 patients had greater QT hysteresis than LQT1 and LQTS-negative patients (Figure 1D, \( P<0.0022 \)). The area under the ROC curve for identifying LQT2 by hysteresis was 0.825 (95% CI, 0.721 to 0.930; \( P<0.0001 \)), for LQT1 by peak QTc shortening was 0.775 (95% CI, 0.654 to 0.896; \( P=0.0002 \)), and for LQTS from postural QTc change was 0.666 (95% CI, 0.544 to 0.789; \( P=0.0095 \); Table 2). In the patients with “concealed” LQTS, postural QTc increase was >30 ms in 68% of patients. Exercise QTc prolongation was >60 ms in 94% of concealed LQT1 patients. Hysteresis was >25 ms in 67% of concealed LQT2 patients.

**Table 1. Study Population Characteristics**

<table>
<thead>
<tr>
<th>Gene Negative (n=64)</th>
<th>LQT1 (n=50)</th>
<th>LQT2 (n=45)</th>
<th>( p )</th>
</tr>
</thead>
<tbody>
<tr>
<td>Women, %</td>
<td>61</td>
<td>56</td>
<td>76</td>
</tr>
<tr>
<td>Age, y</td>
<td>27 (27)</td>
<td>26 (31)</td>
<td>26 (32)</td>
</tr>
<tr>
<td>Rest HR, bpm</td>
<td>79 (25)</td>
<td>71 (15)</td>
<td>71 (20)</td>
</tr>
<tr>
<td>Rest QT, ms</td>
<td>380 (45)</td>
<td>430 (50)</td>
<td>440 (70)</td>
</tr>
<tr>
<td>Rest QTc, ms</td>
<td>426 (33)</td>
<td>461 (43)</td>
<td>482 (39)</td>
</tr>
</tbody>
</table>

HR indicates heart rate.

Values are expressed as median (interquartile range).
Effect of β-Blocker Therapy

The effect of β-blockade is summarized in Table 3. Treated patients had significantly lower resting and peak exercise heart rates. Furthermore, β-blockers normalized the postural QTc prolongation and QT hysteresis. Finally, there was a trend toward β-blocker therapy reducing the QTc prolongation observed during treadmill testing in LQT1 patients (from 65 [IQR, 60] to 47 [IQR, 45] ms; \( P = 0.0785 \)).

Discussion

The primary findings of this study of the utility of an exercise protocol to identify patients with LQT1 and LQT2 mutations were (1) LQTS patients had a greater prolongation of their QTc interval with changes in posture than control subjects; (2) patients with LQT1 mutations had marked QTc prolongation with exercise; (3) LQT2 patients had an exaggerated QT hysteresis compared with LQT1 and control subjects; and (4) β-blockade normalized postural and exercise-induced QTc prolongation and QT hysteresis.

The high prevalence of “concealed” LQTS patients with normal-to-borderline QTc intervals and the high degree of heterogeneity and variable penetrance of LQTS mutations contribute to the ongoing challenge of LQTS diagnosis. A provocative testing strategy to unmask the LQTS phenotype and point to a specific genotype would be of considerable value. In a study of 82 patients with genetically identified LQTS, Takenaka et al showed that a modified Bruce protocol used in combination with a qualitative assessment of T-wave morphology was useful in identifying patients with LQT1 mutations, who experienced marked QTc prolongation during exercise. The identification of LQT2 patients was much more limited in this study as it was dependent on the qualitative assessment of T-wave morphology changes during exercise. In a similar study of 147 genotyped patients, Vyas et al showed that an increase in the QT interval ≥30 ms during epinephrine infusion, a strategy pioneered independently by Shimizu and Ackerman, could identify LQT1 patients with a good degree of accuracy. Furthermore, in a study of 103 patients, Swan et al suggested that a steep QT/R-R slope may be useful in identifying patients with LQT2 mutations. Finally, our preliminary observations suggested that postural changes in QTc and prolonged QT hysteresis during exercise testing may be helpful in identifying patients with LQTS. The current study suggests that failed QT shortening with exercise is characteristic of LQT1, and exaggerated hysteresis is characteristic of LQT2. Combining postural changes with QT/R-R findings from exercise testing is useful in identifying LQTS patients, predicts genotype in a large proportion of patients, and may be useful in directing genetic testing.

In this study, we found that approximately 44% of patients had a “concealed” QTc interval consistent with previous data from Priori et al, highlighting the low penetrance of LQTS and diagnostic challenge these patients can present. Further-
and vagally mediated pathways to sustain the appropriate response involving sympathetic repolarization. Furthermore, sudden changes in posture lead to an exaggerated QT difference between the exercise and recovery QT/R-R curves that is manifested as increased QT hysteresis. This consequently leads to an exaggerated QT difference between the exercise and recovery QT/R-R curves that is manifested as increased QT hysteresis.

In agreement with previous studies, we have also observed that LQT1 patients had an attenuated QT shortening and an exaggerated QTc prolongation during exercise testing. The LQT1 gene encodes for the IKs potassium channel, which is responsible for the repolarization phase of the cardiac cycle at rapid heart rates. In the absence of functional IKs, consistent with what we observed in our LQT1 patient cohort, the QT fails to adapt (ie, shorten) with increasing heart rate. On the other hand, we have also observed that patients with LQT2 mutations have normal QT shortening and minimal prolongation of their QTc interval during exercise. This latter observation is in agreement with previous data from Swan et al,21 who proposed that LQT2 patients could be identified by their steeper QT/HR slope during exercise.

A unique aspect of the current study is the identification of exaggerated QT hysteresis as an LQT2 phenomenon (Figure 2B). ROC curve analysis identified QT hysteresis as a good predictor of LQT2 phenotype. Before the era of genetic testing, we reported exaggerated RT hysteresis as a characteristic of LQTs based on a Schwartz score alone.2,15 In retrospect, the modest-sized study population was subsequently found to be composed primarily of LQT2 patients. Our data thus extend these previous observations and suggest that increased QT hysteresis may be a phenomenon unique to LQT2 patients. QT hysteresis is normally measured at 1 to 2 minutes into the recovery phase, when heart rates typically return to approximately 100 bpm. In LQT2 patients with impaired IKs, the QT fails to shorten at these intermediate heart rates in early exercise, a so-called "IKs zone." This is followed by recruitment of IKs above 100 bpm through to peak exercise with concomitant appropriate QT shortening, which persists into the recovery phase. This consequently leads to an exaggerated QT difference between the exercise and recovery QT/R-R curves that is manifested as increased QT hysteresis.

In agreement with our preliminary observations,12,22 β-blockade normalized QT hysteresis and the QTc prolongation seen with assuming the standing position. In addition,
Atenolol -blockers appeared to attenuate the QTc prolongation seen with exercise. These findings suggest that β-blockade improves QT adaptation to changes in heart rate, perhaps via a direct effect on the “abruptness” of heart rate changes, or an indirect effect on catecholamines or potassium ion channels. Furthermore, as a result of these changes, testing of suspected LQTS patients while on β-blockers may mask the LQTS phenotype during exercise and postural testing, limiting test utility. We did not directly address the optimal means to determine β-blocker efficacy, but we typically target a reduction in peak treadmill heart rate of 30 bpm at peak work load as a practical target for β-blocker effect.

Several forms of exercise were assessed, with consistent changes noted to distinguish LQTS and genotype. Because findings were consistent, we recommend gradual supine bicycle testing because signal artifact from upper body motion during exercise is minimized.

This study has several limitations. First, our observations are based on a modest sample of patients and may not apply to all genotypes of LQTS. Second, in determining the QT interval, identifying the end of the T-wave can be challenging, especially at rapid heart rates. QT hysteresis is determined by calculating the difference between QT intervals during recovery and exercise phases and is typically small; any errors in measuring the QT segment may underestimate or overestimate the true hysteresis value. Third, the Bazett formula was used for correction of the QT interval and may be inaccurate at high heart rates. Because the same correction was applied to all groups, this should not have influenced the study results substantively. Finally, LQTS mutations vary in severity, depending on the location and nature of the mutation. We analyzed mutations by affected gene and, given the small sample size, did not perform a subgroup analysis by mutation severity. Finally, a validation cohort would have strengthened the findings, which we are currently collecting data on. Despite these limitations, the current study found clear utility in combining posture and exercise parameters in identifying and predicting genotype in suspected LQTS patients. Further validation with a larger cohort seems appropriate.

In conclusion, QT hysteresis and QTc prolongation with standing and during exercise is useful in the identification of patients with LQTS. β-Blocker therapy normalized these parameters. Postural and exercise testing is an efficient, accessible, and simple test that is a valuable adjunct to genetic testing.

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**Disclosures**

None.

**References**


![Figure 2. Effect of posture on QTc and QT hysteresis. A, ECGs showing QTc prolongation with postural change in an LQT1 patient. B, ECGs showing QT hysteresis in an LQT2 patient.](http://circep.ahajournals.org/Downloadedfrom)
Exercise Testing to Identify Long-QT Syndrome

Long QT syndrome (LQTS) is a leading cause of sudden death in children and adolescents. Excellent outcomes in the management of LQTS can be achieved with lifestyle modifications and therapeutic intervention, highlighting the importance of its identification. Although genetic testing remains the diagnostic “gold standard,” the diagnosis of LQTS remains challenging when electrocardiographic findings are borderline, which can occur in up to 50% of cases. In this article, we sought to develop a provocative exercise testing strategy to unmask the LQT phenotype and help guide genetic testing. We found that QTc prolongation when comparing lying with standing at the beginning of exercise testing was useful in identifying LQTs patients [P=0.029]. Comparing exercise with recovery QT intervals, increased QT hysteresis was suggestive of LQT2 genotype [P=0.0022]. Failed QT shortening at peak exercise identified LQT1, similar to previous reports. The area under the ROC curve for identifying LQTS from postural QTc change was 0.666 (P=0.0095), and for predicting LQT2 from hysteresis, it was 0.825 (P<0.0001). Postural QTc increase was >30 ms in 68% of “concealed” LQTs patients, and QT hysteresis was >25 ms in 67% of concealed LQT2 patients. A simple postural test in combination with comparison of exercise and recovery QTs is useful in identifying LQTS and predicting genotype. These findings may be useful in directing genetic testing.
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