Utility of the Ventricular Fibrillation Waveform to Predict a Return of Spontaneous Circulation and Distinguish Acute From Post Myocardial Infarction or Normal Swine in Ventricular Fibrillation Cardiac Arrest

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Background—In cardiac arrest, the ventricular fibrillation (VF) waveform, particularly amplitude spectral area (AMSA) and slope, predicts the return of spontaneous circulation (ROSC), but it is unknown whether the predictive utility differs in an acute myocardial infarction (MI), prior MI, or normal myocardium and if the waveform can distinguish the underlying myocardial state. We hypothesized that in a swine model of VF cardiac arrest, AMSA and slope predict ROSC after a shock independent of substrate and distinguish an acute from nonacute MI state.

Methods and Results—MI was induced by occlusion of the left anterior descending artery. Post MI swine recovered for a 2-week period before induction of VF. VF was untreated for 8 minutes in 10 acute MI, 10 post MI, and 10 control swine. AMSA and slope predicted ROSC after a shock independent of myocardial state. For AMSA >31 mV-Hz, the odds ratio was 62 (*P*≤0.001) compared with AMSA <19 mV-Hz. For slope >3.1 mV/s, odds ratio was 52 (*P*≤0.001) compared with slope <1.8 mV/s. With chest compressions, AMSA and slope were significantly lower for acute MI swine compared with control swine, whereas in post MI swine the waveform characteristics were similar to control swine. In particular, for an AMSA >33.5 mV-Hz, the sensitivity to identify an acute from nonacute (control or post MI) state was 83%.

Conclusions—In a swine model of VF cardiac arrest, AMSA and slope predict ROSC independent of myocardial substrate. Furthermore, with chest compressions, the VF waveform evolves differently and may offer a means to distinguish an acute MI. (***Circ Arrhythm Electrophysiol.*** 2011;4:337-343.)

Key Words: cardiopulmonary resuscitation ■ myocardial infarction ■ heart arrest ■ ventricular fibrillation ■ defibrillation

Out-of-hospital cardiac arrest is estimated at 295 000 cases per year in the United States, with 23% caused by ventricular fibrillation (VF) or other initially shockable rhythm.[1,2] In a study of patients ages 50 to 79 years,[3] two-thirds of cardiac arrest patients had preexisting heart disease, with one-third having a prior myocardial infarction (MI). In an autopsy study of sudden cardiac death victims, an active coronary lesion (active thrombus and/or disrupted plaque) was seen in 57%, with acute MI identified by the presence of coagulation necrosis of myocytes, present in 21%[4] and healed, nonacute MI present in 41%. Patients under the age of 50 years have also been demonstrated to have extensive coronary artery disease on autopsy.[5] Furthermore, acute MI with acute coronary occlusion is present in survivors but may not be evident on a postresuscitation ECG,[6,7] which has prompted the recommendation that survivors of cardiac arrest be immediately taken to the cardiac catheterization laboratory.[8]

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It is unclear if the optimal timing of defibrillation and chest compressions is altered in the presence of an acute MI or preexisting coronary disease or heart failure. We have previously shown in a swine study that survival with good neurological outcome is improved by delivering defibrillation first, before the onset of chest compressions in acute myocardial ischemia[9] but not in normal myocardium.[10] For VF of short duration, <4 minutes, the myocardium is physiologically regarded as being in the "electric phase," in which defibrillation is highly likely to restore a perfusing rhythm and therefore the optimal intervention is immediate shock.[11] In a swine study of VF of short duration, the optimal intervention was defibrillation in either an acute MI or normal myocardium.[12] When untreated VF is prolonged beyond 4 minutes and up to about 10 minutes, the myocardium is described as being in the circulatory phase,[11] and in this phase the characteristics of the VF waveform, including parameters based on frequency and amplitude such as amplitude spectral area (AMSA) and slope, have shown good promise as predictors for achieving a return of spontaneous circulation (ROSC) in human and animal studies.[13–21] However, it is unclear whether the predictive ability of AMSA and slope to achieve ROSC is affected by the underlying myocardial substrate, such as the

presence of an acute MI, a prior MI without acute ischemia, or normal myocardium. Furthermore, survival of cardiac arrest hinges on not only care during resuscitation but also care in the postresuscitation period, including emergent cardiac catheterization for acute MI. Because the postresuscitation ECG may not reflect an acute coronary occlusion, other means to identify these patients are needed, in particular if immediate coronary catheterization resources are not available. The VF waveform itself may also help guide this assessment and the triage of patients to emergent cardiac catheterization.

The present study examines the effect of acute or prior MI on the prediction of ROSC based on waveform and whether the waveform can itself reflect the underlying myocardial substrate. In a swine study of VF cardiac arrest we investigate 3 underlying myocardial substrates, an acute MI with acute occlusion of the left anterior descending (LAD) artery, a prior, nonacute, MI, and normal noninfarcted myocardium. We hypothesized that regardless of underlying myocardial substrate that AMSA and slope would predict ROSC and that the evolution of AMSA and slope during resuscitation would distinguish an acute MI from a nonacute MI state.

Methods

Experimental protocols follow the guidelines of the American Physiological Society and were approved by the University of Arizona Institutional Animal Care and Use Committee. Anesthesia and instrumentation were performed by a protocol that is standard in our laboratory. In brief, anesthesia was induced using 5% isoflurane in 100% oxygen delivered by a nose cone, followed by endotracheal intubation. Mechanical ventilation used a mixture of room air and titrated isoflurane (1.5% to 3%), at 12 respirations per minutes and tidal volume of 15 mL/kg, adjusted to keep the end-tidal pCO_2 at 40 ± 3 mm Hg. Anesthesia was kept to the lowest concentration that prevented animal movement and was discontinued before the induction of ventricular fibrillation. Millar (MP-500) microtip, solid-state pressure transducers were used for measurement of aortic and right atrial pressures, and a fluid-filled Swan-Ganz catheter was used for measurements of cardiac output. Correct catheter placement was verified by fluoroscopy. Measurements of ECG, aortic pressure, and right atrial pressure were continuously recorded to a Ponemah Physiology Platform (Data Sciences International, St Paul, MN). External defibrillation patches were placed right parasternal and left lateral, and the signal from the patches was continuously recorded to the defibrillator (Medtronic LifePak 12, Redmond, WA), with a recording sample rate of 125 samples per second. After each experiment, data from the defibrillator were downloaded to a desktop computer for further analysis.

An acute MI was induced by placement of a stainless steel plug in the mid LAD artery, using an Amplatz coronary catheter to engage the left coronary artery. Occlusion of the vessel was confirmed by angiography with contrast injection. Anesthesia was kept to the lowest concentration to prevent animal movement. This method is standard in our laboratory and typically results in infarction of the distal anterior and apical myocardium. For animals in the acute MI group, if VF did not occur spontaneously by 15 minutes after plug placement or if the pig developed severe hypotension, VF was then induced by application of a 100-Hz alternating current through a pacing catheter in the right ventricle. Electric induction of VF after a waiting period of 15 minutes in acute MI animals was chosen to avoid large variability in total ischemic time before initiation of resuscitation. Animals in the control group underwent electric induction of VF after baseline hemodynamic parameters were obtained. Animals in the acute MI and control groups were also used in a recently reported study.[12] Animals in the post MI group were given lidocaine (2 mg/kg) during plug placement to avoid VF during the acute MI. After a 2-week recovery period, the animal was returned to the laboratory and hemodynamic measurements (aortic, right atrial, pulmonary artery, and pulmonary capillary wedge pressure) were made, followed by electric induction of VF with a 100-Hz alternating current delivered from a pacing catheter in the right ventricle. At the end of the experiment, the animal was euthanized and the heart was examined to confirm the presence of scar.

After 8 minutes of untreated VF for swine in all groups, resuscitation commenced with a defibrillation shock of 150 J of biphasic energy. Mechanical ventilation was resumed at a rate of 12 respirations per minute and tidal volume of 15 mL/kg, as used before the induction of VF. Manual chest compressions were provided at a metronome-guided rate of 100 compressions per minute if a perfusing rhythm was not immediately obtained after the shock, and ventilation with oxygen was commenced at a rate of 10/min. ROSC was defined as an unassisted pulse with a peak systolic aortic pressure >50 mm Hg and a pulse pressure of at least 20 mm Hg, lasting for at least 1 minute. After 2 minutes of chest compressions, rhythm was reanalyzed with a ten second "hands-off" interval to allow for an artifact-free period for the off-line data analysis of the VF waveform, described below. Two minutes of uninterrupted chest compressions followed each defibrillation shock and was continued if there was pulseless electric activity. Resuscitation was continued until ROSC was obtained, or efforts were terminated if ROSC was not achieved by 25 minutes. ROSC was assigned to a particular defibrillation shock if it occurred by the end of the 2-minute postshock chest compression interval. Resuscitation time was defined as time from the first defibrillation shock. All shocks were of 150 J. Epinephrine (0.02 mg/kg) was administered intravenously immediately after the first shock if a perfusing rhythm was not obtained and repeated every 3 minutes until ROSC was obtained, for a total of 3 doses.

Data Analysis

The VF signal immediately preceding each defibrillation shock, taken from the 10-second hands-off interval, was analyzed in an 8.2-second interval to determine AMSA and slope, as previously described.[12] AMSA was calculated as the summed product of frequency and square root of power at that frequency, from 4 to 48 Hz, as computed from a fast Fourier transform. AMSA essentially reflects the product of mean frequency and amplitude. Slope was computed from the median of the absolute value of differences in signal voltage every 10 ms in the same 8.2-second interval before a shock. Slope can be thought of as reflecting the product of median frequency and overall amplitude. Thus AMSA and slope measure very similar information from the VF waveform, one from the frequency domain (AMSA) and one from the time domain (slope).

Statistics

Data are presented as mean±SD. A Kruskal-Wallis rank test was performed for comparisons of baseline weight and hemodynamics among the 3 groups of swine.

Logistic regression was performed to determine factors that predicted ROSC, and factors tested were AMSA, slope, MI state (acute, post MI, and control), and resuscitation time, starting with the first shock. In multivariate analyses, AMSA and slope were not tested together because these parameters are highly collinear because they measure very similar information from the VF waveform. In this investigation, the correlation coefficient between AMSA and slope was 0.95. Analysis of factors for the prediction of ROSC included all shocks. The cut-points for slope and AMSA were based on tertiles, which were prespecified. This choice was made to allow for the potential nonlinearity in its association with the outcome of ROSC. The number of bins (tertiles) was chosen so that there would be an ample (of order at least 20) data points within each bin. Logistic regression was also used to assess whether AMSA or slope could predict an acute MI versus nonacute (control or post MI) status. Differences in AMSA and slope among the 3 groups of swine (control, acute MI, and post MI), were assessed by linear regression. Generalized linear mixed models were used to account for correlations within an individual animal due to multiple shocks (ie, multiple measurements of the waveform), using random effects. A receiver operator characteristic (ROC) curve analysis was performed to determine sensitivity and specificity for AMSA and slope to predict

Table 1.   Baseline Characteristics

	Weight, kg	Aortic Systolic Pressure, mm Hg	Right Atrial Pressure, mm Hg	Heart Rate, bpm	Cardiac Output, L/min
Acute MI (n=10)	25.8±2.3	84±13	11±6.6	105±11	2.9±0.6
Post MI (n=10)	27.9±3.0	89±10	11±6.6	108±14	2.7±0.6
Control (n=10)	25.0±2.5	87±9.7	6±3.9	112±14	2.9±2.6

All data given as mean±SD.

ROSC and to predict an acute MI state versus nonacute (control or post MI) state.

Results

A total of 30 swine were studied (10 acute MI, 10 post MI, and 10 control). Baseline characteristics (weight, right atrial pressure, heart rate, systolic aortic pressure, and cardiac output) were similar in all groups of swine before the induction of VF (Table 1).

VF occurred spontaneously in 3 acute MI swine and was induced electrically in the remaining 7, with VF induced in 1 animal at 6 minutes after plug placement because of hypotension. A total of 33 shocks were delivered to acute MI swine, 25 shocks to post MI swine, and 18 shocks to control swine. All control swine achieved ROSC by the second shock, and 2 control swine achieved ROSC with the first shock alone. Two acute MI swine did not achieve ROSC. VF recurred after ROSC had been initially achieved in 2 acute MI swine: 1 at 27 minutes post ROSC, and another animal at 15 minutes and again at 25 minutes. One acute MI animal achieved ROSC within 2 minutes of the first shock. All post MI swine achieved ROSC by the third shock. One post MI animal refibrillated after first achieving ROSC but achieved ROSC again after an additional 10 minutes of resuscitation.

The time course of the waveform characteristics during resuscitation including outcome of each shock for each animal in each group is shown in Figure 1 for AMSA and Figure 2 for slope. AMSA and slope were predictive of ROSC after a shock (Table 2). The highest tertile of AMSA (>31 mV-Hz) had an odds ratio for achieving ROSC of 62 (95% confidence interval [CI], 7.0 to 550; $P<0.001$) compared with the lowest tertile (<19 mV-Hz), with the middle tertile (19 to 31 mV-Hz) having an odds ratio of 10 (95% CI, 1.2 to 89; $P=0.04$). The area under the ROC curve was 0.88. For an AMSA of 32 mV-Hz, the sensitivity to predict ROSC was 68%, with a specificity of 88%, positive predictive value of 76%, and negative predictive value of 82%. The highest tertile of slope (>3.1 mV/s) had an odds ratio for achieving ROSC of 52 (95% CI, 5.9 to 450; $P<0.001$) compared with the lowest tertile (slope <1.8 mV/s). The middle tertile of slope (from 1.8 to 3.1 mV/s) had an odds ratio for achieving ROSC of 12 (95% CI, 1.4 to 105; $P=0.02$) compared with the lowest tertile of slope. An ROC analysis gave an area under the curve of 0.89, and for a slope of 3.1 mV/s, the sensitivity to predict ROSC was 68%, with a specificity of 83%, positive predictive value of 70%, and negative predictive value of 82%.
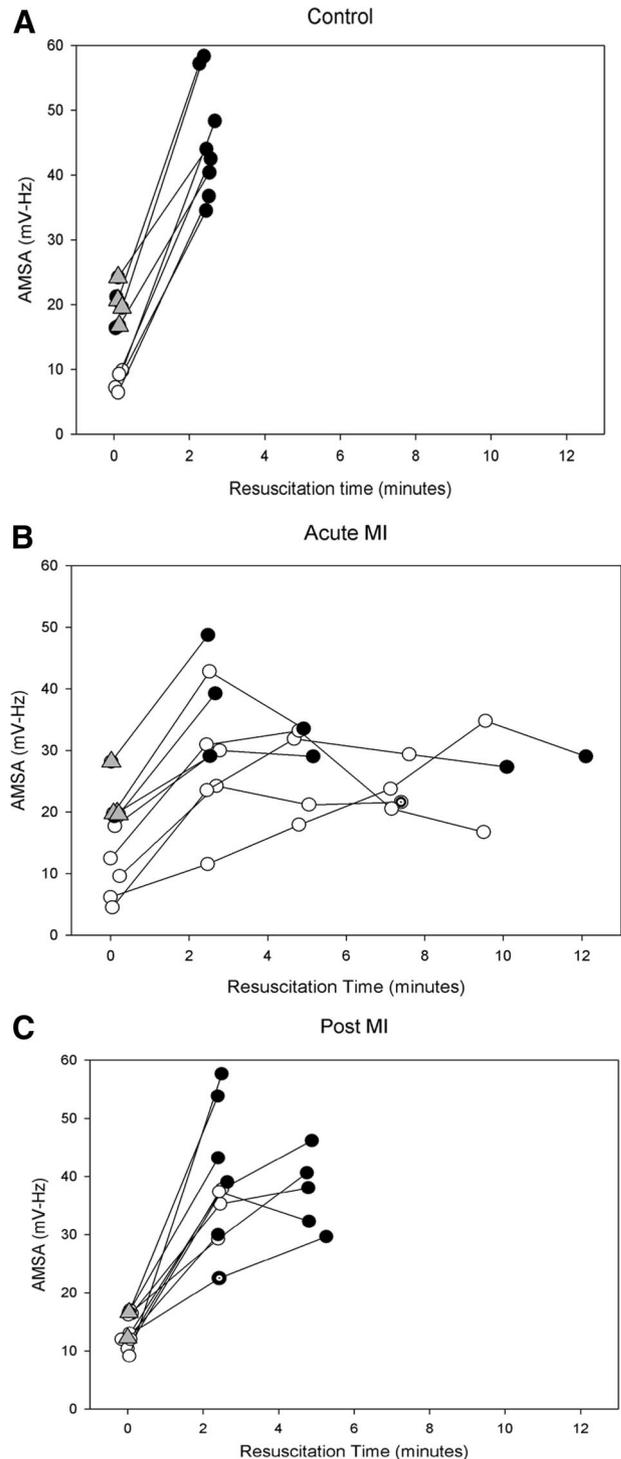
Figure 1. Evolution of AMSA over resuscitation time for control (**A**), acute MI (**B**), and post MI (**C**) swine. The outcome of each shock for each animal (n=10 for each group shown as a connecting line for each animal) is shown by type of marker: open circle indicates VF; black filled circle, ROSC; gray triangle, pulseless electric activity followed by refibrillation; and black circle with inner white dot, transient perfusing rhythm without ROSC followed by refibrillation.

Resuscitation time was not predictive of ROSC (P=NS). MI status was predictive for achieving ROSC: Acute MI swine had an odds ratio of 0.26 (95% CI, 0.08 to 0.87; P=0.03) compared with control swine, and post MI swine
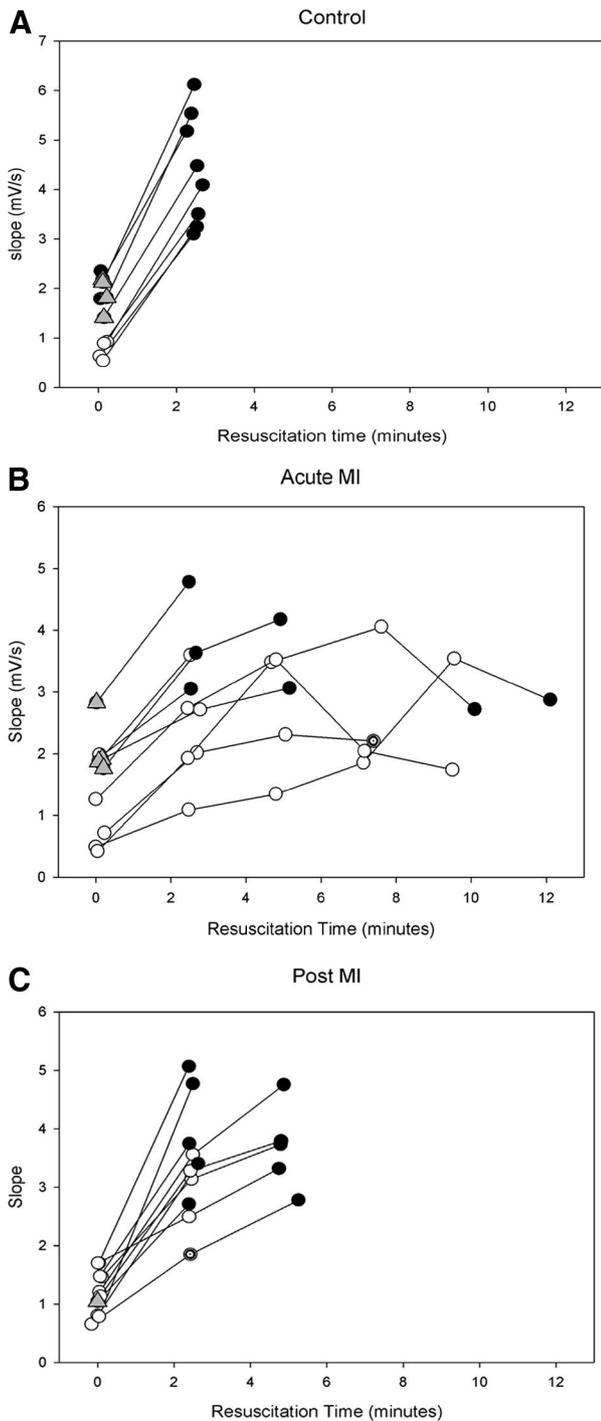
A

B

C

Figure 2. Evolution of slope over resuscitation time for control (**A**), acute MI (**B**), and post MI (**C**) swine. The outcome of each shock for each animal (n=10 for each group shown as a connecting line for each animal) is shown by type of marker: open circle indicates VF; black filled circle, ROSC; gray triangle, pulseless electric activity followed by refibrillation; and black circle with inner white dot, transient perfusing rhythm without ROSC followed by refibrillation.

Table 2. Univariate Logistic Regression Analysis for the Prediction of ROSC

Factor	Odds Ratio	95% CI	*P* Value
AMSA			
19 to 31 mV-Hz*	10	1.2–89	0.04
>31 mV-Hz*	62	7.0–550	<0.001
Slope			
1.8 to 3.1 mV/s*	12	1.4–105	0.02
>3.1 mV/s*	52	5.9–450	<0.001
MI status			
Acute MI†	0.26	0.08–0.87	0.03
Post MI†			NS
Resuscitation time			NS

*Compared with lowest tertile.
†Compared with control swine.

was highly significant as an independent predictor of ROSC, with $P<0.001$, and the middle tertile of slope, with $P=0.009$. For MI status compared with AMSA, the presence of an acute MI also remained a weak but significant independent predictor ($P=0.04$). The highest tertile of AMSA was highly significant as an independent predictor of ROSC, with $P<0.001$, with the middle tertile significant, with $P=0.015$.

Waveform characteristics measured before the first shock were similar between acute MI, post MI swine, and control swine (Figure 3). Before the first shock, acute MI swine had an AMSA of 15.7 ± 7.3 mV-Hz compared with 13.5 ± 2.9 mV-Hz for post MI swine and 15.1 ± 6.4 mV-Hz for control swine ($P=NS$). Before the first shock, slope was 1.5 ± 0.8 mV/s for acute MI swine compared with 1.2 ± 0.4 for post MI swine and 1.5 ± 0.7 mV/s for control swine ($P=NS$). However, after the initiation of chest compressions, AMSA was larger for post MI and control compared with acute MI animals (Figure 3). Before second or higher shocks, AMSA was significantly lower for acute MI swine, 28 ± 9 mV-Hz, compared with control swine, 45 ± 9 mV-Hz ($P=0.001$), whereas AMSA in post MI swine, 38 ± 9 mV-Hz, was similar to control swine ($P=0.18$ for post MI compared with control). Before second and higher shocks, slope was 2.8 ± 1.0 mV/s for acute MI swine compared with 4.4 ± 1.1 mV/s for control swine ($P=0.002$ for acute MI compared with control), whereas for post MI swine, slope was 3.5 ± 0.9 mV/s ($P=0.06$ for post MI compared with control).

Waveform characteristics were also predictive of whether an acute MI was present compared with a nonacute MI (post MI or control) state, after the initiation of resuscitation (measured before second or later shocks). AMSA was predictive of whether an acute MI was present ($P=0.001$), with an area under the ROC curve of 0.85. For AMSA >33.5 mV-Hz, the sensitivity to identify an acute MI state (fraction of shocks from acute MI swine correctly classified as coming from acute MI swine) was 83%, with a specificity (fraction of shocks from nonacute MI swine that are correctly classified as coming from nonacute MI swine) of 78%, with a positive predictive value of 79% and negative predictive value of 82%. Slope was also predictive for the presence of an acute MI versus a nonacute MI (control or post MI) state, $P=0.006$, with area under the ROC of 0.75. For slope >3.3 mV/s, the sensitivity to identify an acute MI state
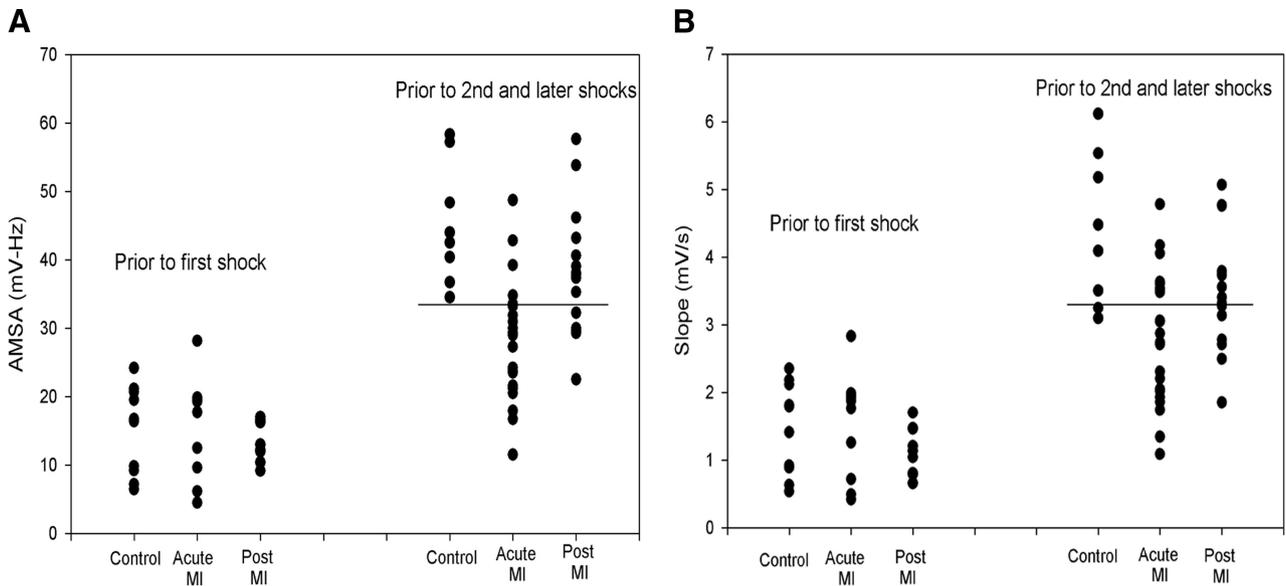
had an odds ratio of 0.53 (95% CI, 0.16 to 1.8; $P=NS$) for achieving ROSC compared with control swine. In a multivariable analysis of MI status compared with slope, the presence of acute MI remained a weak but significant independent predictor ($P=0.02$). The highest tertile of slope

Figure 3. AMSA (**A**) and slope (**B**) for first compared with second or later shocks. For untreated VF before the first shock, AMSA and slope are similar between acute, post MI, and control swine. After the initiation of resuscitation, AMSA measured before second or later shocks increases to a greater extent for control animals compared with acute MI swine (*P*=0.001), whereas post MI swine increased in AMSA to a similar extent as control swine (*P*=0.18). Similarly, slope increased with second or later shocks to a greater extent for control animals compared with acute MI swine (*P*=0.002), and for post MI swine, the change in slope was similar to control swine (*P*=0.06). Both AMSA and slope could distinguish an acute from a nonacute MI (post MI or control) state. For AMSA >33.5 mV-Hz (horizontal line), the sensitivity to identify an acute MI state (fraction of shocks from acute MI swine correctly classified as coming from acute MI swine) was 83%. For slope >3.3 mV/s (horizontal line), the sensitivity to identify an acute MI state was 65%.

was 65%, with a specificity of 65%, positive predictive value of 65%, and negative predictive value of 65%.

Discussion

The present investigation finds that in a swine model of VF cardiac arrest, AMSA and slope are predictive of achieving ROSC regardless of myocardial state, but the waveform characteristics evolve differently in acute MI once resuscitation (ie, chest compressions) is commenced. In untreated VF, waveform characteristics were similar in acute MI, post MI, and control swine. However, after the initiation of resuscitation, namely chest compressions, AMSA and slope were significantly lower in acute MI swine, and both AMSA and slope could distinguish an acute MI status. For AMSA >33.5 mV-Hz, the sensitivity to correctly identify an acute MI was 83%, and for a slope >3.3mV/s, the sensitivity to correctly identify an acute MI was 65%.

Differences in the evolution of the VF waveform according to myocardial state may be related to alterations in the underlying electrophysiological substrate. Optical mapping studies have demonstrated that VF is maintained by the formation of rotors that propagate wave fronts that fragment in surrounding tissues creating successive wavelets, referred to as fibrillatory conduction.[22] Alterations in sodium and potassium membrane currents, as can occur in ischemic and nonischemic myocardium, can modify how rotors form and how fibrillatory conduction proceeds.[22–24] Dominant frequency and number of singularity points have been studied by optical mapping in isolated hearts and shown to be slower in ischemia and heart failure.[25]

Characteristics of the VF waveform have been studied for their ability to predict ROSC after a shock in both human[13–16] and animal[17–21] studies, but with variable reported predictive power. As a result, it has remained unclear if waveform

parameters are sufficiently useful as a threshold to decide whether to deliver a shock or continue chest compressions. There is further uncertainty as to whether different thresholds may apply, based on the underlying myocardial substrate.

We have previously demonstrated in a swine model of VF arrest that AMSA and slope predict ROSC in second and later shocks regardless of the presence or absence of an acute MI, compared with normal swine.[12] A more clinically relevant comparison is the inclusion of preexisting coronary disease with prior MI, which is modeled in the present study. We find that in a multivariable analysis of VF waveform and myocardial status that the VF waveform, AMSA, or slope remains highly predictive (*P*<0.001 for highest tertiles of AMSA or slope) of achieving ROSC, whereas myocardial status (acute MI versus control) is weakly predictive of the outcome of a shock. The presence of a post MI status compared with control was not predictive of ROSC after a shock. Our work implies that the VF waveform is the most important factor to account for in an algorithm to decide when to shock.

Nonetheless, it has been demonstrated in human and swine studies that VF waveform parameters are affected by myocardial substrate and by chest compressions. In swine studies during untreated VF, VF frequency characteristics had an altered time course in different myocardial substrates (acute MI, post MI, and normal myocardium), whereas AMSA and slope were similar among these substrates.[26,27] Treatment with chest compressions, however, does alter the VF waveform, as studied in normal swine[28] and human studies.[29] After the initiation of chest compressions in swine in which VF was induced by balloon occlusion of the LAD, animals with the deflated catheter remaining in place in the LAD had a lower value of AMSA compared with animals in which the balloon catheter was withdrawn.[16]

A differential effect of chest compressions (or other resuscitation interventions), depending on myocardial status, may explain the results reported from an analysis of cardiac arrest victims in Norway, where median slope and AMSA were depressed in patients with an acute MI compared with nonacute MI states, although in that study there was considerable overlap in waveform between patients with and those without an acute MI.[30] We find in this swine study that although AMSA and slope are similar in untreated VF in control, acute, and post MI, they evolve differently once resuscitation, in particular, the delivery of high-quality chest compressions, is commenced. Although overlap in the VF waveform remains between myocardial substrates (Figure 3), we have found that after the initiation of resuscitation, an AMSA >33.5 mV-Hz has a sensitivity of 83% to detect an acute MI from nonacute (control or post MI) state and for slope >3.1 mV/s a sensitivity of 65%. This work would imply that the VF waveform responds differently to well-performed resuscitation interventions according to the underlying myocardial state.

The present study has implications for optimizing postresuscitation care. Both acute and prior nonacute MI are important etiologies for cardiac arrest, as identified both in autopsy data[4] and survivors.[7] In particular, the postresuscitation ECG may not indicate the presence of an acute MI from an acute coronary stenosis. In 435 patients resuscitated from VF cardiac arrest who underwent immediate coronary angiography from the PROCAT registry,[7] a significant coronary lesion was identified in 58% of patients who did not show ST-segment elevation on a postresuscitation ECG and that successful coronary intervention improved hospital survival regardless of the findings on a postresuscitation ECG. In that registry, the most frequent culprit lesion was in the LAD artery. Patients with an acute MI resuscitated from VF arrest are recommended to be taken immediately to the cardiac catheterization laboratory.[8] If laboratory facilities are not immediately available, the patient may require transport to a facility with cardiac catheterization availability. The present study has shown that waveform characteristics respond differently to chest compressions, based on whether an acute or nonacute MI state is present, and suggests that it may be possible to identify patients who are not having an acute MI and who would otherwise not require emergent cardiac catheterization. Along with other markers of acute MI that may or may not be present, such as substantially elevated cardiac enzymes or wall motion abnormalities, as well as the postresuscitation ECG, the waveform evolution during resuscitation may be very useful adjunctive information for the triage of patients to the cardiac catheterization laboratory.

Limitations

There are limitations inherent in animal studies and their applicability to human cardiac arrest. In particular, resuscitation in this model is performed with high-quality chest compressions with minimal interruptions, whereas in human cardiac arrest, chest compressions are performed by lay personnel only 20% to 40% of the time[2,31,32] or with a shallow depth and long interruptions for other interventions such as endotracheal intubation by emergency personnel,[33,34] and for which survival is substantially improved if such interruptions

in chest compressions are avoided.[32] Therefore, the usefulness of examining the change in the VF waveform to predict myocardial state in human arrest may be lost if chest compressions are not performed well. In the present study, epinephrine is given within the first minute of the onset of chest compressions, and we have not addressed which of these resuscitation interventions, compressions or epinephrine, are of greatest importance for the differential changes in VF waveform, based on myocardial state. Further animal and human studies are needed to determine if changes in VF waveform can predict myocardial state in a prospective fashion and whether decisions to shock or continue compressions, based on the VF waveform, can result in an earlier restoration of a perfusion rhythm and improved survival.

We also induced VF in acute MI animals electrically if it did not occur spontaneously by 15 minutes after placement of the plug in the LAD to avoid large variation in total ischemic time. One study[35] has suggested that resuscitation outcome, including number of shocks and time to achieve ROSC, is different in acute MI animals that achieve VF spontaneously versus electrically. In our study, 3 animals in the acute MI group achieved VF spontaneously within 15 minutes of ischemic time, and we cannot assess whether the method of achieving VF is a factor in either resuscitation outcome or change in VF waveform.

Conclusions

In a swine model of cardiac arrest caused by VF, we find that AMSA and slope predict the ROSC after the initiation of resuscitation interventions including chest compressions, regardless of the presence of an acute or prior MI or normal myocardium. AMSA and slope evolve differently in these myocardial substrates and may offer a method to distinguish an acute MI.

Disclosures

None.

References

1. Lloyd-Jones D, Adams RJ, Brown TM, Carnethon M, Dai S, De Simone G, Ferguson TB, Ford E, Furie K, Gillespie C, Go A, Greenlund K, Haase N, Hailpern S, Ho PM, Howard V, Kissela B, Kittner S, Lackland D, Lisabeth L, Marelli A, McDermott MM, Meigs J, Mozaffarian D, Mussolino M, Nichol G, Roger VL, Rosamond W, Sacco R, Sorlie P, Thom T, Wasserthiel-Smoller S, Wong ND, Wylie-Rosett J. Heart disease and stroke statistics: 2010 update: a report from the American Heart Association. *Circulation*. 2010;121:e46–e215.
2. Nichol G, Thomas E, Callaway CW, Hedges J, Powell JL, Aufderheide TP, Rea T, Lowe R, Brown T, Dreyer J, Davis D, Idris A, Stiell I. Regional variation in out-of-hospital cardiac arrest incidence and outcome. *JAMA*. 2008;300:1423–1431.
3. Rea TD, Pearce RM, Raghunathan TE, Lemaitre RN, Sotoodehnia N, Jouven X, Siscovick DS. Incidence of out-of-hospital cardiac arrest. *Am J Cardiol*. 2004;93:1455–1460.
4. Farb A, Tang AL, Burke AP, Sessums L, Liang Y, Virmani R. Sudden coronary death: frequency of active coronary lesions, inactive coronary lesions, and myocardial infarction. *Circulation*. 1995;92:1701–1709.
5. Schmermund A, Schwartz RS, Adamzik M, Sangiorgi G, Pfeifer EA, Rumberger JA, Burke AP, Farb A, Virmani R. Coronary atherosclerosis in unheralded sudden coronary death under age 50: histo-pathologic comparison with 'healthy' subjects dying out of hospital. *Atherosclerosis*. 2001;155:499–508.

6. Spaulding CM, Joly LM, Rosenberg A, Monchi M, Weber SN, Dhainaut JF, Carli P. Immediate coronary angiography in survivors of out-of-hospital cardiac arrest. *N Engl J Med*. 1997;336:1629–1633.

7. Dumas F, Cariou A, Manzo-Silberman S, Grimaldi D, Vivien B, Rosencher J, Empana JP, Carli P, Mira JP, Jouven X, Spaulding C. Immediate percutaneous coronary intervention is associated with better survival after out-of-hospital cardiac arrest: insights from the PROCAT (Parisian Region Out of Hospital Cardiac Arrest) registry. *Circ Cardiovasc Interv*. 2010;3:200–207.

8. Bobrow BJ, Kern KB. Regionalization of postcardiac arrest care. *Curr Opin Crit Care*. 2009;15:221–227.

9. Indik JH, Hilwig RW, Zuercher M, Kern KB, Berg MD, Berg RA. Preshock cardiopulmonary resuscitation worsens outcome from circulatory phase ventricular fibrillation with acute coronary artery obstruction in swine. *Circ Arrhythm Electrophysiol*. 2009;2:179–184.

10. Berg RA, Hilwig RW, Ewy GA, Kern KB. Precountershock cardiopulmonary resuscitation improves initial response to defibrillation from prolonged ventricular fibrillation: a randomized, controlled swine study. *Crit Care Med*. 2004;32:1352–1357.

11. Weisfeldt ML, Becker LB. Resuscitation after cardiac arrest: a 3-phase time-sensitive model. *JAMA*. 2002;288:3035–3038.

12. Indik JH, Allen D, Shanmugasundaram M, Zuercher M, Hilwig RW, Berg RA, Kern KB. Predictors of resuscitation in a swine model of ischemic and nonischemic ventricular fibrillation cardiac arrest: superiority of amplitude spectral area and slope to predict a return of spontaneous circulation when resuscitation efforts are prolonged. *Crit Care Med*. 2010;38:2352–2357.

13. Ristagno G, Gullo A, Berlot G, Lucangelo U, Geheb E, Bisera J. Prediction of successful defibrillation in human victims of out-of-hospital cardiac arrest: a retrospective electrocardiographic analysis. *Anaesth Intensive Care*. 2008;36:46–50.

14. Neurauter A, Eftestol T, Kramer-Johansen J, Abella BS, Sunde K, Wenzel V, Lindner KH, Eilevstjonn J, Myklebust H, Steen PA, Strohmenger HU. Prediction of countershock success using single features from multiple ventricular fibrillation frequency bands and feature combinations using neural networks. *Resuscitation*. 2007;73:253–263.

15. Lin LY, Lo MT, Ko PC, Lin C, Chiang WC, Liu YB, Hu K, Lin JL, Chen WJ, Ma MH. Detrended fluctuation analysis predicts successful defibrillation for out-of-hospital ventricular fibrillation cardiac arrest. *Resuscitation*. 2010; 81:297–301.

16. Ristagno G, Tang W, Xu TY, Sun S, Weil MH. Outcomes of CPR in the presence of partial occlusion of left anterior descending coronary artery. *Resuscitation*. 2007;75:357–365.

17. Nowak CN, Fischer G, Neurauter A, Wieser L, Strohmenger HU. Prediction of countershock success: a comparison of autoregressive and fast Fourier transformed spectral estimators. *Methods Inf Med*. 2009;48: 486–492.

18. Povoas HP, Weil MH, Tang W, Bisera J, Klouche K, Barbatsis A. Predicting the success of defibrillation by electrocardiographic analysis. *Resuscitation*. 2002;53:77–82.

19. Marn-Pernat A, Weil MH, Tang W, Pernat A, Bisera J. Optimizing timing of ventricular defibrillation. *Crit Care Med*. 2001;29:2360–2365.

20. Menegazzi JJ, Callaway CW, Sherman LD, Hostler DP, Wang HE, Fertig KC, Logue ES. Ventricular fibrillation scaling exponent can guide timing of defibrillation and other therapies. *Circulation*. 2004;109:926–931.

21. Menegazzi JJ, Wang HE, Lightfoot CB, Fertig KC, Chengelis NL, Sherman LD, Callaway CW. Immediate defibrillation versus interventions first in a swine model of prolonged ventricular fibrillation. *Resuscitation*. 2003;59:261–270.

22. Vaquero M, Calvo D, Jalife J. Cardiac fibrillation: from ion channels to rotors in the human heart. *Heart Rhythm*. 2008;5:872–879.

23. Zaitsev AV, Guha PK, Sarmast F, Kolli A, Berenfeld O, Pertsov AM, de Groot JR, Coronel R, Jalife J. Wavebreak formation during ventricular fibrillation in the isolated, regionally ischemic pig heart. *Circ Res*. 2003; 92:546–553.

24. Wit AL, Dillon SM, Coromilas J, Saltman AE, Waldecker B. Anisotropic reentry in the epicardial border zone of myocardial infarcts. *Ann N Y Acad Sci*. 1990;591:86–108.

25. Moreno J, Zaitsev AV, Warren M, Berenfeld O, Kalifa J, Lucca E, Mironov S, Guha P, Jalife J. Effect of remodelling, stretch and ischaemia on ventricular fibrillation frequency and dynamics in a heart failure model. *Cardiovasc Res*. 2005;65:158–166.

26. Indik JH, Donnerstein RL, Berg RA, Hilwig RW, Berg MD, Kern KB. Ventricular fibrillation frequency characteristics are altered in acute myocardial infarction. *Crit Care Med*. 2007;35:1133–1138.

27. Indik JH, Donnerstein RL, Hilwig RW, Zuercher M, Feigelman J, Kern KB, Berg MD, Berg RA. The influence of myocardial substrate on ventricular fibrillation waveform: a swine model of acute and postmyocardial infarction. *Crit Care Med*. 2008;36:2136–2142.

28. Berg RA, Hilwig RW, Kern KB, Ewy GA. Precountershock cardiopulmonary resuscitation improves ventricular fibrillation median frequency and myocardial readiness for successful defibrillation from prolonged ventricular fibrillation: a randomized, controlled swine study. *Ann Emerg Med*. 2002;40:563–570.

29. Eftestol T, Wik L, Sunde K, Steen PA. Effects of cardiopulmonary resuscitation on predictors of ventricular fibrillation defibrillation success during out-of-hospital cardiac arrest. *Circulation*. 2004;110:10–15.

30. Olasveengen TM, Eftestol T, Gundersen K, Wik L, Sunde K. Acute ischemic heart disease alters ventricular fibrillation waveform characteristics in out-of hospital cardiac arrest. *Resuscitation*. 2009;80:412–417.

31. Aufderheide TP, Yannopoulos D, Lick CJ, Myers B, Romig LA, Stothert JC, Barnard J, Vartanian L, Pilgrim AJ, Benditt DG. Implementing the 2005 American Heart Association guidelines improves outcomes after out-of-hospital cardiac arrest. *Heart Rhythm*. 2010;7:1363–1364.

32. Bobrow BJ, Clark LL, Ewy GA, Chikani V, Sanders AB, Berg RA, Richman PB, Kern KB. Minimally interrupted cardiac resuscitation by emergency medical services for out-of-hospital cardiac arrest. *JAMA*. 2008;299:1158–1165.

33. Wik L, Kramer-Johansen J, Myklebust H, Sorebo H, Svensson L, Fellows B, Steen PA. Quality of cardiopulmonary resuscitation during out-of-hospital cardiac arrest. *JAMA*. 2005;293:299–304.

34. Valenzuela TD, Kern KB, Clark LL, Berg RA, Berg MD, Berg DD, Hilwig RW, Otto CW, Newburn D, Ewy GA. Interruptions of chest compressions during emergency medical systems resuscitation. *Circulation*. 2005;112: 1259–1265.

35. Niemann JT, Rosborough JP, Youngquist S, Thomas J, Lewis RJ. Is all ventricular fibrillation the same? A comparison of ischemically induced with electrically induced ventricular fibrillation in a porcine cardiac arrest and resuscitation model. *Crit Care Med*. 2007;35:1356–1361.

CLINICAL PERSPECTIVE

Ventricular fibrillation and pulseless ventricular tachycardia are the initial rhythm in about one-quarter of cardiac arrests. Although the performance of effective chest compressions with minimal to no interruptions is well known to be a critical component in resuscitation, the optimal duration of chest compressions before defibrillation shocks is unclear. It is unknown if the timing of shocks and chest compressions should be the same for patients with acute myocardial infarction (MI) as those with a history of MI, or even those without any cardiovascular disease. Furthermore, it is recognized that patients with acute MI resuscitated from cardiac arrest caused by ventricular fibrillation may not show ST-segment elevation, complicating decisions of whether to send patients for emergent cardiac catheterization once a perfusing rhythm has been restored. This investigation explores the evolution of the ventricular fibrillation waveform parameters of amplitude spectral area and slope in acute MI, post MI, and control swine. We find that amplitude spectral area and slope predict the restoration of a perfusing rhythm after a shock independent of the underlying myocardium, and furthermore with well-performed chest compressions, amplitude spectral area at a threshold of 33.5 mV-Hz showed a sensitivity of 83% to distinguish an acute from a nonacute MI state.

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Utility of the Ventricular Fibrillation Waveform to Predict a Return of Spontaneous Circulation and Distinguish Acute From Post Myocardial Infarction or Normal Swine in Ventricular Fibrillation Cardiac Arrest

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