Acute and Chronic Performance Evaluation of a Novel Epicardial Pacing Lead Placed by Percutaneous Subxiphoid Approach in a Canine Model

Roy M. John, MBBS, PhD; Kevin Morgan, AA; Lucas H. Brennecke, DVM; Michael E. Benser, PhD; Pierre Jais, MD

Background—Endovascularly implanted leads risk vascular injury and endocarditis, and can be difficult to locate in desired positions for LV pacing. We evaluated the acute and long-term stability, electric performance and histopathology of a percutaneously placed intrapericardial lead (IPL).

Methods and Results—Twelve adult mongrel dogs underwent defibrillator implants incorporating IPLs. Successful uncomplicated percutaneous implantation of an IPL was achieved in all. Early fluoroscopic shift noted with 3 of 6 of the initial version IPL-1 was not seen with the modified IPL-2. Mean±95% confidence interval bipolar capture threshold at 0.5-ms pulse width for the IPL increased from 0.69±0.14 V at implant to 1.50±0.34 V (P=0.003) at 12 weeks. The 12-week thresholds were higher for IPL compared with right ventricular endocardial leads (0.75±0.33 V; P=0.001) but not different compared with coronary sinus leads (1.33±0.58 V; P=0.994). IPL impedance increased from 742±46 Ω at implant to 1066±207 Ω at 12 weeks (P=0.007). R-wave amplitude at 12 weeks was 8.37±1.52 mV. There was no important phrenic nerve stimulation from IPL pacing. Histopathology in 8 animals showed adequate adhesion of the electrodes or mesh to the epicardium without damage to underlying vasculature. There was no evidence for late pericardial inflammation or effusion.

Conclusions—The IPL demonstrated adequate stability of position and acceptable electric parameters without chronic pericardial inflammation in this canine model and offers a potential alternative to endocardial pacing leads. (Circ Arrhythm Electrophysiol. 2015;8:659-666. DOI: 10.1161/CIRCEP.114.002076.)

Key Words: cardiac pacemaker, artificial • epicardial pacing • percutaneous epicardial pacing • pericardium
WHAT IS KNOWN

- Endovascularly placed pacing and defibrillating leads risk vascular injury, endocarditis and extracardiac stimulation, and maybe difficult to position in desired locations for left ventricular pacing.
- Access to the pericardium can be safely obtained percutaneously.

WHAT THE STUDY ADDS

- In a systematic evaluation of a novel percutaneously introduced intrapericardial lead, adequate pacing and sensing thresholds comparable with endocardially implanted coronary sinus leads were demonstrated in 12 canine experiments.
- Modifications to the original lead design improved stability within the pericardial space with adequate fixation to the epicardium. There was no evidence for chronic pericardial inflammation, effusion, or damage to the epicardial surface or underlying blood vessels.
- Intrapericardial lead has potential applications in cardiac pacing when transvenous access is not possible or desirable.

Endovascularly placed pacing and defibrillating leads risk vascular injury, endocarditis and extracardiac stimulation, and maybe difficult to position in desired locations for left ventricular pacing. Access to the pericardium can be safely obtained percutaneously.

12 adult mongrel dogs weighing 20 to 26 kg were implanted with an implantable cardioverter defibrillator with cardiac resynchronization therapy under general anesthesia. Using sterile technique with the animal in a supine position, a subxiphoid incision was made to the deep fascia and puncture of the pericardium was performed under fluoroscopy using a Tuohy needle, aided by injection of contrast and use of a guidewire to confirm entry to the space. A dedicated short steerable sheath (Agilis PF Introducer, St. Jude Medical) was used to deliver 1 IPL to the pericardial space and for positioning to overlie the lateral left ventricular epicardium, where satisfactory electric performance was confirmed and registered. During the procedure, care was taken to prevent air from entering the pericardial space. If necessary, air was removed using the side port available on the Agilis PF Introducer and a syringe. Using the introducer and syringe in this manner also allowed to check for bleeding within the pericardial space. In an attempt to prevent lead migration, suture sleeves with 3 ligatures were used to affix the lead to subcutaneous tissue at the subxiphoid location. The lead was then tunneled submuscularly to an incision at the last rib space and then subcutaneously to a subcutaneous pocket for the pulse generator, which was fashioned with blunt dissection in the left dorsal side of the thorax area. The incision at the subxiphoid access site was closed and sutured.

A standard cut down of the left external jugular vein was performed. Under fluoroscopic guidance, one 7 Fr defibrillation lead with bipolar sensing tip electrodes (St. Jude Medical Models Riata 1580, 1581, or 7001) was placed in the right atrium. The remaining 6 animals received 1 bipolar lead (St. Jude Medical Model 1688T) was placed in the right atrium. The remaining 6 animals received 1 bipolar lead (St. Jude Medical Model 1058T) and 1 active fixation lead (St. Jude Medical Model 1688T) in the right atrium. The IPL was plugged into the LV port and the right atrium or CS lead was plugged into the right atrium port of the device. The leads and device were implanted in the subcutaneous pocket and the pocket was closed using standard surgical procedures.

Figure 1. Ventral view (left) and a schematic representation of a cross-section at the electrode (right) of the intrapericardial lead version 2 (IPL-2). Right, 1 = A layer of insulation overlying the outer surface of the electrode to prevent phrenic nerve capture. 2 = Polyester mesh surrounding the electrode. 3 = Active electrode to contact with the epicardial surface. Note the central bore in the electrode for steroid (not incorporated in the IPLs tested in this study). 4 = Weld of the conductor coil to the ring of the electrode. 5 = Central lumen of the lead.
Intraoperative Testing

Standard measurement of all lead parameters including cathodal pacing thresholds (measured at 0.5 ms pulse width), pacing impedances, and intrinsic amplitudes were made in the bipolar mode. For IPL-2, measurements were also made in the unipolar mode to the RV coil. Phrenic nerve stimulation (PNS) was assessed while pacing via the IPL in bipolar mode up to a maximal output of 7.5 V at 0.5 ms. Fluoroscopic imaging in the anterior–posterior and lateral views, were stored to document the initial position of the IPL. With the animal in a lateral position, the stability of the IPL was challenged with the vigorous skeletal muscle contractions associated with 2 high energy R-wave synchronized shocks at 20 and 30 J delivered successively. After these shocks, repeat fluoroscopy images were obtained to record and any shift in position of the IPL.

Follow-Up

After the implantation procedure and testing, the animals were recovered and maintained in a standard animal housing facility. Electric parameters of all leads that included bipolar cathodal pacing thresholds (at 0.5-ms pulse duration), impedances, and intrinsic amplitudes were recorded at weekly intervals for 6 weeks, 2 weekly intervals to 12 weeks, at 4 weekly intervals to week 104, and thereafter, every 6 weeks to the latest time point or to the point of euthanization. At each pacing check, fluoroscopy images along the same 4 images recorded intraoperatively were collected for offline analysis. In addition, PNS was assessed at each pacing check through visual observation coupled with palpation of the diaphragm during testing. Finally, 2 synchronized programmer shocks as during the intraoperative testing, were delivered at week 4 and 12.

Explants and Pathological Examination

Out of the 12 canines, planned explant and pathological studies were performed in 5 animals at weeks 12, 12, 28, 40, and 104. Two animals were euthanized at weeks 152 and 176 for methicillin-resistant staphylococcal infections, 1 involving the pericardium and another one due to cardiac tamponade or arrhythmias. All 12 implants were achieved without complications with placement of the IPL in the desired location to overlie the left ventricular epicardial surface, 6 with the initial IPL-1 design and 6 subsequently with the IPL-2 design. Procedure time (from when the Agilis P1 Introducer was positioned within the intrapericardial space until the final positioning and electrical testing) for placement of all IPLs was 14 (12–22) minutes.

Stability of IPL

None of the IPL-1 nor IPL-2 leads were judged to have dislocated on delivery of the intraoperative defibrillation shocks. However, 3 of the 6 IPL-1 leads were identified having shifted implant position by fluoroscopic imaging comparison over the follow-up period; 2 were noted to have shifted at 1-week postimplant, and the other at 2 weeks postimplant but without significant changes in electric parameters. After this initial observation, all IPL-1 leads remained stable in subsequent follow up. All six IPL-2 leads remained stable, with no identifiable shift in lead position over the duration of the follow-up (Figure 2).

Electric Performance

The bipolar pacing impedances, capture thresholds, and R-wave amplitudes over the course of follow up of 158 weeks for all the IPLs are shown in Figure 3. The impedance and capture thresholds from the IPLs, taken together, exhibited significant increases over the 12-week maturation period (F test $P<0.001$ for the effect of implant duration on each of impedance and threshold). The IPLs sensing amplitudes did not exhibit a significant change over the 12-week maturation period (F test $P=0.053$). Bipolar capture threshold at 0.5-ms pulse width for all IPLs was 0.69±0.14 V at implant and increased to 1.50±0.34 V at 12 weeks ($t$ test $P=0.003$). Pacing lead impedance for all IPL leads was 742±46 $\Omega$ at implant and increased to 1066±207 $\Omega$ at 12 weeks ($t$ test $P=0.007$). R-wave amplitude at implant was 12.6±3.1 mV and decreased to 8.4±1.5 mV ($t$ test $P=0.034$). There was no difference in the electric performance of the IPL-1 versus the IPL-2 during 12 weeks in implantation, the period over which all animals endured. To address non-normal data, a square root transformation was applied to all lead parameters. Changes in IPL-1 and IPL-2 parameters were taken together over these 12 weeks and were assessed using $F$ tests from a repeated measures model assuming compound symmetry of the correlation matrix. To assess a few judiciously selected time points at which IPL parameters might have changed, paired $t$ tests were used to compare IPL parameters at implant versus 1, 4, and 12 weeks postimplant. Differences in lead parameters between the IPL-1 and IPL-2 leads were assessed using $F$ tests associated with the lead main effect from the repeated measures model.

Results

All 12 implants were achieved without complications with placement of the IPL in the desired location to overlie the left ventricular epicardial surface, 6 with the initial IPL-1 design and 6 subsequently with the IPL-2 design. Procedure time (from when the Agilis P1 Introducer was positioned within the intrapericardial space until the final positioning and electrical testing) for placement of all IPLs was 14 (12–22) minutes. No acute complications occurred during lead placement. In particular, no animal had significant pericardial bleeding leading to cardiac tamponade or arrhythmias.

Pathology

Gross examination of the pericardium, heart surface, and IPL was conducted on each heart. The pericardium was removed, where possible, to determine the degree of pericardial/epicardial adhesions and to expose the IPL. Gross photographs were taken to document the findings. The IPL including tissues adjacent to the 2 electrodes and the polyester mesh were dissected free of the remainder of the heart, and were processed, infiltrated, and embedded in methyl methacrylate. After methyl methacrylate polymerization, radiographs were obtained and were processed, infiltrated, and embedded in methyl methacrylate. The polyester mesh were dissected free of the remainder of the heart. The condition of the pericardial sac was also noted and maintained in a standard animal housing facility. Electric parameters of all leads that included bipolar cathodal pacing thresholds (at 0.5-ms pulse duration), impedances, and intrinsic amplitudes were recorded at weekly intervals for 6 weeks, 2 weekly intervals to 12 weeks, at 4 weekly intervals to week 104, and thereafter, every 6 weeks to the latest time point or to the point of euthanization. At each pacing check, fluoroscopy images along the same 4 images recorded intraoperatively were collected for offline analysis. In addition, PNS was assessed at each pacing check through visual observation coupled with palpation of the diaphragm during testing. Finally, 2 synchronized programmer shocks as during the intraoperative testing, were delivered at week 4 and 12.

Statistical Analysis

Because of the uneven explant schedule of the animals, all analyses of lead parameters were analyzed only over the initial 12 weeks of
capture threshold, R-wave amplitude and impedance (F tests $P=0.206, 0.778$, and 0.493, respectively).

Comparisons of the IPL-2 pacing parameters to those of the RV and CS LV leads at implant and 1, 4, and 12 weeks are shown in Figure 4. Compared with the endocardial active fixation RV lead, IPL-2 had higher capture thresholds at 1, 4, and 12 weeks. However, there were no differences in the IPL-2 capture thresholds compared with those of the endovascular CS lead at any of the time points. Pacing impedances for the IPL-2 were higher than those of the RV lead at implant, 1, 4, and 12 weeks, but not different than those of the CS lead. R-wave sensing for the IPL-2 was comparable to the RV lead and significantly higher than that of the CS lead across the observational period.

Pacing and sensing were tested in the unipolar mode for both proximal and distal electrodes to the RV coil for IPL-2. Highest unipolar pacing threshold at 0.5 ms and lowest R wave for either electrode was $1.0\pm 0.6$ V and $14.4\pm 2.9$ mV, respectively at 12 weeks. Unipolar lead impedances for distal and proximal electrodes were $502\pm 100$ and $542\pm 48$ Ω, respectively at 12 weeks.

None of the 12 animals exhibited PNS intraoperatively, at the stimulus strength of 7.5 V at 0.5 ms. Over the course of follow-up, 11 of the 12 animals did not exhibit any instances of PNS. One animal, implanted with IPL-1, exhibited PNS at pacing amplitudes between 6.0 and 7.5 V during 6 of its 40 follow-up evaluations.

Gross Pathology and Histology
Pathological data were available on all 8 euthanized animals at time periods ranging between 12 and 188 weeks (Table I in the Data Supplement). Figure 5 shows a gross appearance of the

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**Figure 2.** Anterior–posterior fluoroscopic views at implant and at 116 weeks of an intrapericardial lead version 2 (IPL-2) lead implant. The epicardial lead overlies the posterior lateral left ventricle with no discernible change in position during the period of monitoring. CS indicates coronary sinus; and RV, right ventricle.

**Figure 3.** Bipolar pacing capture threshold, R-wave sensing amplitude and pacing impedance for the intrapericardial lead (IPL) leads. Each $P$ value versus implant (t tests). The data represent 12 animals through 12 weeks; 10 through 28; 9 through 40; 8 through 104; 7 through 152; and 6 through 158. Data represented as mean±95% confidence interval.
posterior surface of an explant at 104 weeks. The entire lead including the wing and electrodes could be visualized in this specimen with minimal pericardial thickening or fibrosis. In gross examination, none of the animals developed a pericardial effusion.

There was variable degrees of adhesion and thickening of the pericardium with mild inflammatory changes seen in the early explants. However, there was no evidence of inflammation in the planned explants beyond 40 weeks. In all histological studies, there was good adhesion of the electrodes or mesh with the epicardium without damage to underlying blood vessels (Figure 6). No major epicardial vessels were directly beneath the pacing electrodes or mesh. In 1 IPL-1 explant, there was 3 mm of epicardial fat separating the proximal electrode from the epicardium (Figure 7). Despite this separation, there was adequate pacing threshold with this lead. The distal electrode in this explant was firmly opposed to the epicardium allowing for excellent pacing and sensing thresholds in the bipolar mode. Aside from this, 1 electrode overlying 3-mm epicardial fat, the thickness of fibrotic encapsulation around the electrodes separating them from the epicardium in the IPL-1 explants was 1 mm for 1 electrode, 0.4 to 0.5 mm for 3 electrodes, and <0.1 mm for the remaining electrodes. For the 2 IPL-2 explants, the encapsulation thicknesses around the 4 electrodes were all ≤0.1 mm.

In the explants after euthanization for methicillin-resistant Staphylococcus aureus infection, there was dense pericardial thickening and adhesions to the epicardium. However, there was no evidence for pericardial effusion.

**Discussion**

This report represents the first systematic evaluation of a passive fixation lead system designed for easy percutaneous delivery to the epicardial surface. The present series of experiments provide data on 2 versions of the epicardial

![Figure 5. Gross appearance of an explanted heart at 104 weeks. The posterior view of the heart is shown. The lead with the electrodes, stabilizing wings can be visualized through a translucent pericardium that demonstrated minimal thickening.](image-url)
bipolar pacing lead. The main findings are as follows: (1) epicardial placement of the IPL can be safely achieved in canine models using a subxiphoid approach and dedicated tools to direct placement in areas where adequate pacing and sensing thresholds are obtained; (2) although IPL-1 lead design was associated with early dislodgement within the pericardial space, subsequent modifications in the IPL-2 design provided for lead stability, (3) IPL pacing thresholds and impedance were comparable to endovascular CS leads for LV pacing, but higher than the endocardial RV leads. Sensing for the IPL-2 was similar to that of the RV leads, (4) gross and histological examination of planned and unplanned explants showed early inflammatory changes but free of the development of effusion and adequate fixation of the electrodes to the epicardial surface without damage to myocardium or blood vessels.

Despite the greater ease of transvenous implants, incidence of cardiac implantable electronic device infections with endocarditis has risen over the years with higher rates for patient on dialysis. In addition, chronic upper extremity vascular occlusion and inadequate left ventricular branches to the CS for LV pacing necessitate the use of epicardial pacing wires. Surgical epicardial lead placement is associated with a higher upfront morbidity and mortality. Hence, an IPL that can be introduced percutaneously has appeal. Leads have been placed in pediatric patients by percutaneous subxiphoid pericardial access or via a small surgical subxiphoid incision to access the pericardial space. In the adult population, surgical access requires more extensive dissection. However, percutaneous epicardial access is now commonly used for epicardial ablation of arrhythmias and can be safely achieved in patients without previous cardiac surgery or pericardial disease. In this study using canine models, successful placement of a novel IPL in a desired location in the pericardial space to overlie the posterior lateral LV was successfully achieved in all 12 dogs.

Electric measurements in the IPL lead remained stable over several months after implantation despite the lack of steroid elution in these leads. Although the chronic thresholds in the IPL-2 leads were consistently higher than the endocardial RV active fixation lead, the maximum capture threshold registered was 2.5 V at 0.5 ms and still within the acceptable limits for an epicardial pacing lead. It was comparable to that of the LV leads placed via the CS. Histologically, varying degrees of fibrosis and epicardial fat were present beneath the electrodes. The bipolar design of the IPL leads would be expected to allow successful pacing capture and sensing from at least one of the electrodes.

The histological reaction to endocardial leads is well documented in animals and human studies. Typically, there is thrombus formation, localized inflammatory process with granulation tissue and gradual replacement with fibrosis.
fibrous tissue. Defibrillation leads incite a greater fibrotic reaction compared with pacemaker leads. The findings relating to the passive fixation IPL leads followed a similar pattern but without the initial thrombosis. There was variable fibrotic reaction that resulted in adequate fixation of the electrodes and the surrounding mesh to the epicardial surface or pericardium. The fibrous tissue at the interface seems to be minimal and does not influence the electric measurements over the longer term. No epicardial vessels were entrapped beneath the lead in these experiments but second-order arterioles and venules beneath the electrodes were not affected.

Potential problems relating to an IPL lead include the issue of access to the pericardial space. In adults, access to the pericardial space for epicardial ablation for ventricular tachycardia was first described by Sosa et al and increasingly used for ablation. Nevertheless, a dry pericardial tap can be associated with complications that include intra-abdominal bleeding, laceration of the liver, injury to the coronary arteries, and cardiac perforation. RV puncture is the most common complication encountered in 4.5% of 215 consecutive cases in a series by Sosa et al. However, RV puncture rarely causes continued bleeding and often seals spontaneously. In the absence of previous pericardial inflammation or cardiac surgery, catheters and sheaths can be moved about freely within the pericardial space to deploy a lead in a desired area. In this series, we chose the posterior lateral LV for lead location because this is the area where activation commonly occurs latest during left bundle branch block and is thus, a good location for pacing for CRT. The issue of extracardiac stimulation, for example, phrenic nerve, is theoretically eliminated by the lead design that uses insulation over the outer surface of the electrodes. In this series, no late migration of the lead was seen. However, migration is possible in the presence of undetected pericardial effusion. Finally, in the event of infection that involves the pericardial space, removal will most likely entail surgical intervention. In the early stages, advancement of a sheath over the lead for retraction may be possible but we have no information about safety of this technique. In the present series, 2 animals developed staphylococcal infection, involving the pericardium in 1 and requiring euthanization at weeks 152 and 176. These were likely because of secondary seeding of the devices from an unidentified primary source.

Limitations
Complete data on the 2 animals is only available to 12 weeks and hence the demonstrated stability and performance of this lead is short term. Planned and unplanned sacrifices at various time points have, however, provided data on histopathology spanning 12 to 188 weeks. Four surviving animals have continued to demonstrate adequate lead performance. Finally, epicardial fat layers may be thicker in the human heart with possible interference with pacing capture although it is unusual to encounter significant adipose tissue over low posterior-lateral LV where IPL were positioned in the present series of experiments.

Conclusions
In this first systematic study of a novel percutaneously introduced IPL in the canine model, we have demonstrated adequate stability of the lead in the pericardial space without dislodgement, adequate pacing and sensing threshold and no damage to the epicardial surface or underlying small blood vessels. An IPL lead has potential applications in cardiac pacing when transvenous access is not possible or desirable.

Disclosures
Dr John received modest honoraria for lectures from St. Jude Medical, Inc, Medtronic, Inc, and Boston Scientific, Inc. K. Morgan and Dr Benser are employees of St. Jude Medical, Inc. Dr Jais is a consultant to Biosense Webster, Inc, and St. Jude Medical, Inc, L.H. Brenneck reports no conflicts.

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SUPPLEMENTAL MATERIAL

Table: Time to explant for each experiment, gross and histological findings at pathological examination.
<table>
<thead>
<tr>
<th>Animal No. and type of lead</th>
<th>Explant time</th>
<th>Reason for Explant</th>
<th>General Gross Appearance</th>
<th>Pericardium Gross</th>
<th>Presence of fluid</th>
<th>Histology (presence of fibrosis and extent)</th>
<th>Inflammation yes/no</th>
<th>Underlying myocardium</th>
<th>Epicardial fat/Vessels</th>
</tr>
</thead>
<tbody>
<tr>
<td>1 IPL-1</td>
<td>12 wks</td>
<td>Planned</td>
<td>Mild epicardial bruising beneath distal electrode</td>
<td>Multifocal areas of fibrinous adhesion between epicardium and pericardium</td>
<td>no</td>
<td>Proximal electrode attached to epicardium by mild to moderate fibrous connective tissue and small areas of loose connective tissue.</td>
<td>Small numbers of lymphocytes and plasma cells. Mild to moderate granulomatous inflammation surrounding mesh on epicardial surface.</td>
<td>Normal</td>
<td>No fat or vessels</td>
</tr>
<tr>
<td>2 IPL-1</td>
<td>12 wks</td>
<td>Planned</td>
<td>Both electrodes firmly attached to epicardium and pericardium</td>
<td>Unremarkable</td>
<td>no</td>
<td>Electrodes firmly compressed against epicardium with thin fibrous capsule around the mesh</td>
<td>Mild inflammatory cell infiltration</td>
<td>Underlying myocardium normal</td>
<td>Epicardial fat present and small arteriole within fat unremarkable</td>
</tr>
<tr>
<td>3 IPL-2</td>
<td>28 wks</td>
<td>Planned</td>
<td>Grossly unremarkable</td>
<td>Minimal fibrosis all around lead body from site of entry into pericardium.</td>
<td>no</td>
<td>Fibrous tissue (&lt;100µm) between electrodes and epicardium, Both electrodes separated</td>
<td>Macrophages and giant cell infiltration beneath mesh that had separated</td>
<td>Normal</td>
<td>Small venule and arterioles beneath within epicardial fat between</td>
</tr>
<tr>
<td>IPL</td>
<td>Weeks</td>
<td>Type</td>
<td>Description</td>
<td>Fibrosis</td>
<td>Inflammation</td>
<td>Myocardium</td>
<td>Notes</td>
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<tr>
<td>4</td>
<td>40</td>
<td>Planned</td>
<td>Electrodes firmly adherent to epicardium and mesh encapsulated in fibrous tissue.</td>
<td>No</td>
<td>Minimal fibrosis between electrodes and epicardium but mesh encapsulated in fibrous capsule.</td>
<td>Normal</td>
<td>No blood vessels in vicinity of electrodes/mesh</td>
<td></td>
<td></td>
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<tr>
<td>5</td>
<td>104</td>
<td>Planned</td>
<td>Translucent pericardium with no adhesions.</td>
<td>No</td>
<td>No significant fibrotic reaction around electrodes but mesh attached to myocardium with mild fibrosis.</td>
<td>Normal</td>
<td>Proximal electrode separated from myocardium by 3 mm of fat. Small arteriole and vein unremarkable. Distal electrode in direct contact with myocardium with 50µm of fibrosis.</td>
<td></td>
<td></td>
</tr>
<tr>
<td>6</td>
<td>152</td>
<td>Euthanized for MRSA Infection</td>
<td>Epicardium deep red and had extensive mild multifocal pericardial fibrosis. Both electrodes (&lt;)</td>
<td>No</td>
<td>No inflammatory cell infiltrate.</td>
<td>Normal</td>
<td>No fat or vessels beneath</td>
<td></td>
<td></td>
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<tr>
<td></td>
<td>176 wks</td>
<td>Euthanized for MRSA bacteremia</td>
<td>Minimal fibrotic reaction externally but pericardium firmly attached to epicardium over most of the heart</td>
<td>Both electrodes attached by fibrosis to epicardium and pericardium</td>
<td>no</td>
<td>Minimal fibrosis around polyester mesh attaching mesh to epicardial surface.</td>
<td>No inflammation</td>
<td>Myocardium beneath electrodes and mesh was normal.</td>
<td>0.8 mm of fat beneath electrode but no inflammatory infiltrate. Venule 0.5 mm from electrode in fat unremarkable.</td>
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<tr>
<td>7</td>
<td>IPL-1</td>
<td>188 wks</td>
<td>Euthanized for lymphoma</td>
<td>Pericardium adhered to epicardium. Electrodes encapsulated by fibrous tissue but poor attachment to epicardium</td>
<td>no</td>
<td>Mild fibrosis around proximal electrode but none around distal electrode</td>
<td>No</td>
<td>Normal myocardium</td>
<td>Electrodes separated from myocardium by 200 µm of fat and connective tissue. No blood vessels</td>
</tr>
</tbody>
</table>